





PhD Project Proposal

Funder details	
Studentship funded by:	Convergence Science Centre
Project details	
Project title:	Exploiting 3D in vitro models to advance diffusion magnetic resonance imaging (MRI) as a probe of cellular and tumour microstructure
Supervisory team	
Primary Supervisor:	Dr Jessica Winfield
Associate Supervisor(s):	Dr Paula Cunnea (Lead Supervisor 2) Dr Clementine Lesbats Dr Tom Lund Prof Simon Robinson
Secondary Supervisor:	Prof Christina Messiou
Divisional affiliation	
Primary Division:	Division of Radiotherapy and Imaging, The Institute of Cancer Research,

London

Primary Team: Applied Physics in Clinical MRI

Primary Location: Sutton

Project background

There is an unmet clinical need for early detection and characterisation of tumours, residual disease and metastases using non-invasive imaging methods.

Diffusion magnetic resonance imaging (MRI) exploits the motion of water molecules to sensitise the MRI signal to sub-voxel cellular and microstructural properties [Norris 2001, Taouli 2016]. Diffusion MRI is an essential component of clinical MRI in oncology for lesion detection and response assessment including quantitative analysis via the apparent diffusion coefficient (ADC) [Winfield 2019a]. ADC relates to tumour cell density but is also influenced by other factors including the extracellular matrix (ECM) and necrosis, leading to a lack of specificity [Winfield 2019b, Winfield 2021, Reeves 2023]. The lack of specificity impairs the ability of diffusion MRI to distinguish, for example, cellular tumour from fibrosis in stromal dense tumours [Muraoka 2008] or detect tumour infiltrating immune cells in some tumour types [Surov 2023]. Clinical studies to elucidate these factors are limited by complexity of the tumour microenvironment, practical constraints on in vivo measurements, spatial heterogeneity, and often lack a

ground-truth. There is a clinical need for more specific diffusion MRI measurements that can inform on cellular/microstructural properties such as cell sizes, cell density, membrane integrity, properties of the ECM and infiltration by tumour cells or immune cells [McLaughlin 2020]. More advanced models of diffusion MRI offer scope to probe these cellular/microstructural properties [Mitra 1995, Fieremans 2010, Panagiotaki 2014, Jiang 2017]. By employing a 'bottom up' approach using controlled systems and a 'ground-truth' from digital pathology we will determine which models are the most suitable for oncological applications.

The aims of this project are to develop experimental diffusion MR methods to probe cellular/microstructural properties using controlled systems (cell suspensions, cell pellets, spheroids, organoids) alongside digital pathology. The project will follow an iterative process of biological development, MR experiment and analysis, pathology and modelling in systems of increasing complexity. The study will test the hypothesis that advanced diffusion MR can non-invasively determine cellular/microstructural properties such as cell sizes, cell density, membrane integrity, ECM properties and infiltration.

Project aims

- Develop experimental diffusion MRI methods in cell suspensions using samples that recapitulate relevant cellular/microstructural properties.
- Evaluate models of the diffusion MRI measurements in cell suspensions and evaluate modelled values against digital pathology measurements.
- Use high-resolution diffusion MRI of cell pellets, spheroids and organoids to evaluate diffusion MRI models against digital pathology measurements.
- Select appropriate diffusion MRI models for clinical applications by simulation of motion of water molecules in samples (cell suspensions, cell pellets, spheroids, organoids) using digital pathology images and evaluate against experimental results.

Research proposal

The team has expertise on MR systems at a wide range of field strengths (1.5T-11.7T) with clinical and pre-clinical measurement capabilities, enabling complementary methods and enhancing translation. The team has expertise in using a variety of cell lines, which are available for the project. Immortalised cell lines and patient-derived organoids representing ovarian high-grade serous cancer will be used as an example cancer system as it is a cancer of unmet need, which displays complex spatial heterogeneity with peritoneal dissemination in advance stages with ECM remodelling and variable immune invasion of tumours [Ploski 2021, Clark 2022, Cunnea 2023].

The project is divided into three work-packages (WP):

WP1. Development of diffusion MRI methods to evaluate cellular/microstructural properties in cell suspensions.

We will produce suspensions of immortalised cell lines in gels suitable for diffusion MRI experiments. Sample preparation will be based on previously reported methods [Lundberg 1994, Katashima 2013] and extended to evaluate cellular/microstructural properties and treatment effects. Samples will recapitulate relevant cellular/microstructural properties to evaluate the influence of cell sizes, cell density, membrane integrity and ECM on diffusion MRI. Physical (heat, ultrasound) and chemical treatments will be used to alter cell membrane integrity. Other agents (collagen, fibronectin, other ECM proteins) will be added to alter ECM composition and structure. Tumour cell lines with altered expression of factors implicated in the ECM will be compared to parental cell lines. Digital pathology will provide 'ground-truth' readouts of cell sizes, cell density and structure/properties of the extracellular space. Diffusion MRI measurements will use a clinical high-field (3T) MRI scanner at RMH to estimate ADC and time-dependent diffusion measurements using short and long diffusion times. Measurements will be extended using ultra-high-field NMR using diffusion-ordered spectra at variable diffusion times and echo times to probe diffusion coefficients and relaxation times of water molecules in different compartments. Models will be fitted to the diffusion MRI measurements and modelled values (e.g. cell density) evaluated against digital pathology measurements.

<u>Outcome</u>: Experimental assessment of diffusion model parameters in cell suspensions and evaluation of models compared with ground-truth digital pathology measurements.

WP2. High-resolution diffusion MRI in cell pellets, spheroids, and organoids.

Samples will be constructed with increasing complexity (cell pellets/spheroids/organoids) to extend methods from WP1 into more clinically-realistic systems. Samples will be grown and embedded in gels or ECM materials for diffusion MRI using methods developed at ICR to enable imaging of single cell pellets/spheroids/organoids. Ultra-high field MRI systems will be used to enable high spatial resolution required (7T pre-clinical MRI scanner at ICR; pan-London 7T clinical MRI scanner where ICR is a partner). Diffusion MRI will be used to estimate ADC and time-dependent diffusion measurements using short and long diffusion times. Initially cell pellets and spheroids grown from single immortalised cell lines will be investigated and extended to organoids and co-cultures of spheroids or organoids with normal and cancer associated fibroblasts/stromal cells and/or immune cells to create more complex/realistic samples. Drug treatments for ovarian cancer will be performed with standard-of-care chemotherapies (carboplatin, paclitaxel) or targeted therapies (PARP inhibitors). Radiotherapy treatments will be administered using a cell irradiator. Samples will be embedded and sectioned for digital pathology analysis aligned with the MR imaging plane to assess cell sizes, cell density and identify other cells/materials present (e.g. fibroblasts, collagen, endothelial cells) and responses to treatment.

<u>Outcome</u>: Experimental assessment of ADC estimates and time-dependent diffusion measurements in cell pellets, spheroids, and organoids, and comparison with ground-truth measurements from digital pathology.

WP3. Simulating motion of water molecules using digital pathology images and comparison with experimental results.

Digital pathology images from WP1-2, including annotations identifying cells and other materials, will be used to simulate motion of water molecules in biological structures, including intra/extracellular contributions, exchange between intra/extracellular compartments and ECM properties. Cell suspensions will be simulated initially and extended to more complicated structures (cell pellets/spheroids/organoids). Simulations will investigate the influence of cell sizes, cell densities and membrane permeability on diffusion MRI properties, and results will be evaluated against experimental results from WP1-2.

<u>Outcome</u>: Selection of best diffusion model following evaluation of simulated and experimental diffusion coefficients in biological structures.

WP1 will be conducted in Year 1, providing a foundation in biological methods (growth of cell lines, production of cell suspensions) and MR methods (acquisition, analysis) and will build foundations in digital pathology.

WP2 will be conducted in Years 2 and 3. The student will develop more advanced tissue culture methods using spheroids and organoids and will develop more challenging MR acquisition methods for diffusion MRI of individual spheroids and organoids.

WP3 will be conducted in Year 3 and part of Year 4. The student will develop a deeper understanding of diffusion MRI models via modelling using digital pathology images.

The student will be based at ICR/RMH (Sutton), where MRI experiments will be undertaken. The student will spend time in Dr Cunnea/Professor Fotopoulou's lab at ICL throughout their studentship.

Literature references

CLARK, J., FOTOPOULOU, C., CUNNEA, P. & KRELL, J. 2022. Novel Ex Vivo Models of Epithelial Ovarian Cancer: The Future of Biomarker and Therapeutic Research. *Front Oncol*, 12, 837233.

CUNNEA, P., CURRY, E. W., CHRISTIE, E. L., NIXON, K., KWOK, C. H., PANDEY, A., WULANDARI, R., THOL, K., PLOSKI, J., MORERA-ALBERT, C., MCQUAID, S., LOZANO-KUEHNE, J., CLARK, J. J., KRELL, J., STRONACH, E. A., MCNEISH, I. A., BOWTELL, D. D. L. & FOTOPOULOU, C. 2023. Spatial and temporal intra-tumoral heterogeneity in advanced HGSOC: Implications for surgical and clinical outcomes. *Cell Rep Med*, 4, 101055.

FIEREMANS, E., NOVIKOV, D. S., JENSEN, J. H. & HELPERN, J. A. 2010. Monte Carlo study of a two-compartment exchange model of diffusion. *NMR Biomed*, 23, 711-24.

JIANG, X., LI, H., XIE, J., MCKINLEY, E. T., ZHAO, P., GORE, J. C. & XU, J. 2017. In vivo imaging of cancer cell size and cellularity using temporal diffusion spectroscopy. *Magn Reson Med*, 78, 156-164.

KATASHIMA, K., KURODA, M., ASHIDA, M., SASAKI, T., TAGUCHI, T., MATSUZAKI, H., MURAKAMI, J., YANAGI, Y., HISATOMI, M., HARA, M., KATO, H., OHMURA, Y., KOBAYASHI, T., KANAZAWA, S., HARADA, S., TAKEMOTO, M., OHNO, S., MIMURA, S. & ASAUMI, J. 2013. In vitro assessment of factors affecting the apparent diffusion coefficient of Jurkat cells using bio-phantoms. *Acta Med Okayama*, 67, 359-67.

LUNDBERG, P., ROY, S. & KUCHEL, P. W. 1994. Immobilization methods for NMR studies of cellular metabolism--a practical guide. *Immunomethods*, 4, 163-78.

MCLAUGHLIN, M., PATIN, E. C., PEDERSEN, M., WILKINS, A., DILLON, M. T., MELCHER, A. A. & HARRINGTON, K. J. 2020. Inflammatory microenvironment remodelling by tumour cells after radiotherapy. *Nat Rev Cancer*, 20, 203-217.

MITRA, P. P., LATOUR, L. L., KLEINBERG, R. L. & SOTAK, C. H. 1995. Pulsed-Field-Gradient Nmr Measurements of Restricted Diffusion and the Return-to-the-Origin Probability. *Journal of Magnetic Resonance Series A*, 114, 47-58.

MURAOKA, N., UEMATSU, H., KIMURA, H., IMAMURA, Y., FUJIWARA, Y., MURAKAMI, M., YAMAGUCHI, A. & ITOH, H. 2008. Apparent diffusion coefficient in pancreatic cancer: characterization and histopathological correlations. *J Magn Reson Imaging*, 27, 1302-8.

NORRIS, D. G. 2001. The effects of microscopic tissue parameters on the diffusion weighted magnetic resonance imaging experiment. *NMR Biomed*, 14, 77-93.

PANAGIOTAKI, E., WALKER-SAMUEL, S., SIOW, B., JOHNSON, S. P., RAJKUMAR, V., PEDLEY, R. B., LYTHGOE, M. F. & ALEXANDER, D. C. 2014. Noninvasive quantification of solid tumor microstructure using VERDICT MRI. *Cancer Res*, 74, 1902-12.

PLOSKI, J., BURGER-RAMOS, M., YANG, Y., CUNNEA, P. & FOTOPOULOU, C. 2021. 793 Patient-derived organoids reflect intra-tumoural heterogeneity in high grade serous ovarian cancer. *International Journal of Gynecologic Cancer*, 31.

REEVES, E. L., LI, J., ZORMPAS-PETRIDIS, K., BOULT, J. K. R., SULLIVAN, J., CUMMINGS, C., BLOUW, B., KANG, D., SINKUS, R., BAMBER, J. C., JAMIN, Y. & ROBINSON, S. P. 2023. Investigating the contribution of hyaluronan to the breast tumour microenvironment using multiparametric MRI and MR elastography. *Mol Oncol*, 17, 1076-1092.

SUROV, A., EGER, K. I., POTRATZ, J., GOTTSCHLING, S., WIENKE, A. & JECHOREK, D. 2023. Apparent diffusion coefficient correlates with different histopathological features in several intrahepatic tumors. *European Radiology*, 33, 5955-5964.

TAOULI, B., BEER, A. J., CHENEVERT, T., COLLINS, D., LEHMAN, C., MATOS, C., PADHANI, A. R., ROSENKRANTZ, A. B., SHUKLA-DAVE, A., SIGMUND, E., TANENBAUM, L., THOENY, H., THOMASSIN-NAGGARA, I., BARBIERI, S., CORCUERA-SOLANO, I., ORTON, M., PARTRIDGE, S. C. & KOH, D. M. 2016. Diffusion-weighted imaging outside the brain: Consensus statement from an ISMRM-sponsored workshop. *J Magn Reson Imaging*, 44, 521-40.

WINFIELD, J. M., MIAH, A. B., STRAUSS, D., THWAY, K., COLLINS, D. J., DESOUZA, N. M., LEACH, M. O., MORGAN, V. A., GILES, S. L., MOSKOVIC, E., HAYES, A., SMITH, M., ZAIDI, S. H., HENDERSON, D. & MESSIOU, C. 2019a. Utility of Multi-Parametric Quantitative Magnetic Resonance Imaging for Characterization and Radiotherapy Response Assessment in Soft-Tissue Sarcomas and Correlation With Histopathology. *Front Oncol*, 9, 280.

WINFIELD, J. M., WAKEFIELD, J. C., BRENTON, J. D., ABDULJABBAR, K., SAVIO, A., FREEMAN, S., PACE, E., LUTCHMAN-SINGH, K., VROOBEL, K. M. & YUAN, Y. 2021. Biomarkers for site-specific response to neoadjuvant chemotherapy in epithelial ovarian cancer: relating MRI changes to tumour cell load and necrosis. *British Journal of Cancer*, 124, 1-8.

WINFIELD, J. M., WAKEFIELD, J. C., DOLLING, D., HALL, M., FREEMAN, S., BRENTON, J. D., LUTCHMAN-SINGH, K., PACE, E., PRIEST, A. N. & QUEST, R. A. 2019b. Diffusion-weighted MRI in Advanced Epithelial Ovarian Cancer: Apparent Diffusion Coefficient as a Response Marker. *Radiology*, 293, 374-383.

Candidate profile		
Pre-requisite qualifications of applicants:	First- or Upper Second- class Honours degree (or a Masters) in physics, engineering, chemistry, or a related subject.	
Intended learning outcomes:	Training in inter-disciplinary research in basic science with clear scope for clinical translation.	
	Practical skills in tissue culture, including advanced tissue culture methods using spheroids and organoids.	
	Practical skills in acquisition and analysis of diffusion MRI data using clinical and preclinical MRI systems.	
	Data analysis and mathematical modelling.	
	Understanding of digital pathology.	
	Training in statistics, scientific writing, and presentation skills.	
	Understanding of cutting edge biological and physics methods working at the interface of biology and physics.	
Advertising details		
Project suitable for a student with a background in:	 □ Biological Sciences ☑ Physics or Engineering ☑ Chemistry □ Maths, Statistics or Epidemiology □ Computer Science 	